

## Quick-Neuron™ Dopaminergic - Maintenance Medium

Catalog Number: DA-MM

### Introduction

Quick-Neuron™ Dopaminergic - Maintenance Medium may be used for the long-term maintenance of human pluripotent stem cell-derived dopaminergic neurons following differentiation as outlined in the Quick-Neuron™ Dopaminergic - SeV Kit, mRNA Kit, and Human iPSC-derived Neurons user guides. Quick-Neuron™ Dopaminergic differentiated cell cultures display typical neurite outgrowth and express a variety of neuronal markers, such as the pan-neuronal marker tubulin beta 3 class III (TUBB3) and the dopaminergic markers tyrosine hydroxylase (TH) and dopamine (DA). When handled and maintained according to the instructions in this user guide, dopaminergic neurons are viable long-term and are suitable for a variety of characterization and neurotoxicity assays.

**Scale:** The Quick-Neuron™ Dopaminergic - Maintenance Medium provides sufficient medium for 4 wells of a 24-well plate for up to 2 weeks.

**Related Products:** Quick-Neuron™ Dopaminergic - SeV Kit, Catalog Number: DA-SeV  
 Quick-Neuron™ Dopaminergic - mRNA Kit, Catalog Number: DA-mRNA  
 Quick-Neuron™ Dopaminergic - Human iPSC-derived Neurons, Catalog Number: DA-SeV-CW

### Kit Contents

Upon receipt, store the reagents at the temperatures indicated in the table below. All reagents are shipped on dry ice.

Reagents	Volume	Storage
Component N	840 µl	-20°C or -80°C
Component D4	20 µl	-20°C or -80°C
Component D6	16 µl	-20°C or -80°C
Component P	14 µl	-20°C or -80°C

### Required Consumables

Item	Vendor	Catalog Number
DMEM/F12	ThermoFisher	21331020
Neurobasal Medium	ThermoFisher	21103049
Glutamax (100x)	ThermoFisher	35050061
Penicillin-Streptomycin	ThermoFisher	15140122

### Conditions of Use

This product is for research use only. It is not approved for use in humans or for therapeutic or diagnostic use.

### Technical Support

For technical support, please contact us at [cs@elixirgenscientific.com](mailto:cs@elixirgenscientific.com) or call +1 (443) 869-5420 (M-F 9am-5 pm EST).

Last revised: July 14, 2020

## Base Media Preparation

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### Medium N

1. Prepare Medium N using the reagents listed in the table below.
  - Thaw Component N at 4°C overnight or 30 minutes on ice.
  - All other reagents should be warmed at room temperature for 20-30 minutes.

Medium N Reagents	Volume
DMEM/F12	8 ml
Neurobasal Medium	8 ml
200 mM Glutamax (100x)	83 µl
Penicillin-Streptomycin (10000 units/ml; 100x)	167 µl
Component N	517 µl

2. Store Medium N for up to 2 weeks at 4°C.
  - The leftover Component N can be discarded or saved for another use.

### First Week

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#### Medium N(D4D6P)

1. Prepare Medium N(D4D6P) using the reagents listed in the table below.
  - Thaw Component D4 at 4°C overnight or 30 minutes on ice. Spin down before use.
  - All other reagents should be warmed at room temperature for 20-30 minutes. Spin down Component D6 before use.

Medium N(D4D6P) Reagents	Volume
Medium N	5.5 ml
Component D4	5.5 µl
Component D6	5.5 µl
Component P	2.8 µl

2. Save the leftover Component D4 and Component D6 at 4°C.
  - The leftover Component P can be discarded or saved for another use.
3. Warm Medium N(D4D6P) at room temperature for 20-30 minutes until it no longer feels cold.
4. Pipet out half (400 µl) of the old medium from each well using a P1000 pipettor and add 400 µl Medium N(D4D6P).
5. Incubate the cultures at 37°C, 5% CO<sub>2</sub> for 2 days.
6. Repeat Steps 3-5 every 2-3 days such as on Monday, Wednesday, and Friday for 1 week.

## Second Week

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### Medium N(D4D6)

1. Prepare fresh Medium N(D4D6) using the reagents listed in the table below.
  - Place Component D4 on ice. Spin down before use.
  - Warm Component D6 at room temperature for 30 minutes. Spin down before use.
  - Warm Medium N at room temperature for 30 minutes.

Medium N(D4D6) Reagents	Volume
Medium N	7 ml
Component D4	7 $\mu$ l
Component D6	7 $\mu$ l

2. Warm Medium N(D4D6) at room temperature for 20-30 minutes until it no longer feels cold.
3. Pipet out most of the old medium, but not completely (i.e., just enough to cover the surface of the well), from each well using a P1000 pipettor and add 800  $\mu$ l Medium N(D4D6) along the wall of the well very slowly.
4. Incubate the cultures at 37°C, 5% CO<sub>2</sub> for 2 days.
5. For subsequent medium changes, pipet out half (400  $\mu$ l) of the old medium from each well using a P1000 pipettor and add 400  $\mu$ l Medium N(D4D6).
6. Repeat Step 5 every 2-3 days such as on Monday, Wednesday, and Friday for 1 week.